HK36: What is the diagnostic relevance of molecular techniques for polymicrobial periprosthetic joint infections?

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Response/Recommendation:

Molecular diagnostics can enhance the diagnosis of polymicrobial periprosthetic joint infections (PJI). Multiplex PCR (mPCR) and Next Generation Sequencing (NGS) are the two most extensively studied molecular diagnostic technologies in recent years. The average sensitivity of molecular diagnostics for diagnosing polymicrobial PJI is higher than that of conventional microbial culture; however, the average specificity is lower compared to culture. A combination of culture and molecular diagnostics is recommended when polymicrobial PJI is suspected, as originally diagnosing and treating polymicrobial PJIs as monomicrobial is associated with worse outcomes.

Level of Evidence: Limited

Rationale:

To answer the question above (HK36), we conducted a comprehensive literature review across two databases (PubMed and Cochrane Library), utilizing the MeSH terms developed by our librarians. After screening the potentially eligible studies, we included 1,315 studies for full review and data extraction; ultimately, 75 were selected.

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Periprosthetic joint infections (PJI) are considered polymicrobial when at least two different organisms are isolated from synovial fluid or intraoperative tissue cultures (1).

Polymicrobial PJIs account for up to 46% of all PJIs (2) (3). Historically, polymicrobial PJIs have demonstrated a lower success rate than monomicrobial PJIs due to the increased morbidity, costs, and complications usually associated with modern polymicrobial PJI treatments (4). Understanding the microbiological profile of the causative organisms remains central to the treatment strategy, as several species of pathogens are notoriously difficult to eradicate since these pathogens reside in a biofilm rather than in a more metabolically active planktonic form.

There is no consensus on the typical microorganisms responsible for polymicrobial PJIs: Yapar et al. (5) averaged 2.4 microorganisms in their series of polymicrobial PJIs; in the same study, those authors reported *Staphylococcus epidermidis* and *Cutibacterium acnes* (well-known low-virulence bacteria) as the leading pathogens in polymicrobial PJIs. In contrast, Wouthuyzen-Bakker et al. (6), Löwik et al. (7), and Li et al. (1) suggested that additional organisms with relatively low or variable virulence (e.g., *Enterococcus spp.*, anaerobes) or high-virulence organisms (e.g., *Staphylococcus aureus*, gramnegative bacilli) were more often responsible for polymicrobial PJIs.

To counteract the high culture negativity rate (up to 41% in some reports)(8)(9), molecular diagnostic technologies (DNA- and RNA-based) have been recently introduced to the market to quickly identify the infecting microorganisms and their antibiotic resistance (10). Among them, multiplex polymerase chain reaction (mPCR) (11), next-generation sequencing (NGS) (12), including metagenomic NGS (mNGS) (13) (14) (15), showed the best accuracy in identifying polymicrobial PJIs.

The utility of broad-range PCR with Sanger sequencing was lower for polymicrobial infections than that with standard culture or NGS techniques, as the presence of multiple organisms results in overlapping reads that were difficult to interpret (16). Multiplex PCR sensitivity in diagnosing monomicrobial PJI ranged between 33 and 85% (17) (18), while its specificity ranged between 91% and 100% (19) (20); only a few authors (18) (19) supported the diagnostic role of mPCR in polymicrobial PJIs. Lausmann et al. (18) were able to identify additional microorganisms in 12.5% of their intraoperative samples from infected joints analyzed with a new generation mPCR system; Gardete-Hartmann (19) recommended mPCR as an intraoperative test at the time of a 2-stage revision to rule out an incomplete initial diagnosis of complex polymicrobial infection that was not initially captured by culture.

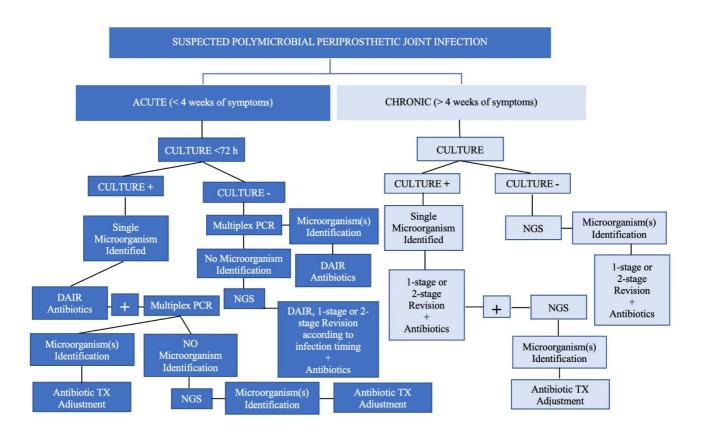
Several authors have compared standard microbial cultures with NGS in systematic reviews and metaanalyses. Kato et al. (21) examined the detection rates of pathogens between NGS and microbial cultures using synovial fluid samples taken from replaced joints; the detection rate for NGS was significantly higher than that for culture (OR 4.52). Different findings emerged from Kildow et al. (22): those authors analyzed samples from 116 patients suspected of having PJI. They demonstrated that NGS did not yield superior sensitivity or specificity results compared to culture. Hantouly et al. (23), in a systematic review and meta-analysis, compared the sensitivity and accuracy of NGS and standard culture: data from 341 patients (seven studies) were reviewed, revealing an overall pooled sensitivity of NGS at 94%, while the pooled specificity was 89%; conversely, the pooled sensitivity of culture was 70%, and the specificity was 94%. In the same study (23), the accuracy of NGS and culture was reported as 91.9% and 80.5%, respectively. Unfortunately, most of these studies (18-20) did not assess the performance of molecular diagnostics in general or NGS specifically for diagnosing polymicrobial PJIs.

Mei et al. (15), in a mid-sized cohort study, were among the few to report that mNGS exhibited high sensitivity (85.7%), but moderate specificity (60%) and accuracy (65.2%) for the diagnosis of polymicrobial PJIs. Interestingly, in that study (15), the specificity, positive predictive value (PPV), and diagnostic accuracy of conventional microbial culture for diagnosing polymicrobial PJI were higher than those of mNGS; in contrast, the sensitivity and negative predictive value (NPV) of conventional microbial culture for diagnosing polymicrobial PJI were lower than those of mNGS. Goswami et al. (24) utilized NGS in a series of culture-negative PJIs: NGS was able to identify a pathogen in 65.9%; in the same study, NGS revealed a polymicrobial PJI in 91% of culture-negative PJIs.

In summary, it is recommended that different testing methods (standard culture and molecular diagnostics) be combined when a polymicrobial PJI is suspected. In clinical practice, the diagnosis of polymicrobial PJI made only by standard culture may be influenced by the concurrent use of antibiotics (25) and the competitive inhibition effects of multiple bacteria during culture (26).

Figure 1 proposes a diagnostic flow chart for the suspect of a polymicrobial PJI.

Figure 1: **Diagnostic flow chart for suspected polymicrobial PJI** when molecular techniques (multiplex PCR; NGS) are available at the treating Institution. The distinction between Acute and Chronic PJI has been made according to the 2018 ICM criteria (27). **Acute PJI**: Debridement, Antibiotics, and Retention of the Implant (DAIR) are recommended only if symptoms persist for less than four weeks with the possible identification of the microorganism; contraindications to DAIR also include the presence of a sinus tract, difficult-to-treat (DTT) infections, loosening of the implant, and antibiotic availability (28) (29). In culture-negative PJI with the presence of a sinus tract, the use of molecular diagnostics must be supported by future research. **Chronic PJI**: contraindications to a 1-stage revision also include the presence of sepsis, an infection caused by drug-resistant bacteria or DTT, the presence of a sinus tract, severe soft-tissue deficiency over the joint, and a history of multiple revisions (30) (31) (4).



References:

- 1. Li H, Fu J, Niu E, Chai W, Xu C, Hao LB, et al. The risk factors of polymicrobial periprosthetic joint infection: a single-center retrospective cohort study. BMC Musculoskelet Disord. 2021 Dec 12;22(1):780.
- 2. Marculescu CE, Cantey RJ. Polymicrobial Prosthetic Joint Infections: Risk Factors and Outcome. Clin Orthop Relat Res. 2008 Jun;466(6):1397–404.
- 3. Moran E, Masters S, Berendt AR, McLardy-Smith P, Byren I, Atkins BL. Guiding empirical antibiotic therapy in orthopaedics: The microbiology of prosthetic joint infection managed by debridement, irrigation and prosthesis retention. Journal of Infection. 2007 Jul;55(1):1–7.
- 4. Palmer JR, Pannu TS, Villa JM, Manrique J, Riesgo AM, Higuera CA. The treatment of periprosthetic joint infection: safety and efficacy of two stage versus one stage exchange arthroplasty. Expert Rev Med Devices. 2020 Mar 3;17(3):245–52.
- 5. Yapar A, Köse Ö, Özdöl Ç, Luo TD, Budin M, Rosa GF, et al. Increased Involvement of Staphylococcus epidermidis in the Rise of Polymicrobial Periprosthetic Joint Infections. J Arthroplasty. 2024 Dec;39(12):3056–61.
- 6. Wouthuyzen-Bakker M. Cultures in periprosthetic joint infections, the imperfect gold standard? EFORT Open Rev. 2023;8(4):175–9.
- 7. Löwik CAM, Zijlstra WP, Knobben BAS, Ploegmakers JJW, Dijkstra B, de Vries AJ, et al. Obese patients have higher rates of polymicrobial and Gram-negative early periprosthetic joint infections of the hip than non-obese patients. PLoS One. 2019 Apr 8;14(4):e0215035.
- 8. Bejon P, Berendt A, Atkins BL, Green N, Parry H, Masters S, et al. Two-stage revision for prosthetic joint infection: predictors of outcome and the role of reimplantation microbiology. Journal of Antimicrobial Chemotherapy. 2011 May 1;66(5):1204–1204.
- 9. Goh GS, Parvizi J. Diagnosis and Treatment of Culture-Negative Periprosthetic Joint Infection. J Arthroplasty. 2022 Aug;37(8):1488–93.
- 10. Gatti G, Taddei F, Brandolini M, Mancini A, Denicolò A, Congestrì F, et al. Molecular Approach for the Laboratory Diagnosis of Periprosthetic Joint Infections. Microorganisms. 2022 Aug 5;10(8).
- 11. Li C, Li H, Yang X, Zhu FZ, Xu C, Trampuz A. Meta-analysis of synovial fluid polymerase chain reaction for diagnosing periprosthetic hip and knee infection. J Orthop Surg Res. 2022 Dec 1;17(1).
- 12. Tang Y, Zhao D, Wang S, Yi Q, Xia Y, Geng B. Diagnostic Value of Next-Generation Sequencing in Periprosthetic Joint Infection: A Systematic Review. Vol. 14, Orthopaedic Surgery. Sociedade Brasileira de Matematica Aplicada e Computacional; 2022. p. 190–8.
- 13. Gu W, Miller S, Chiu CY. Clinical Metagenomic Next-Generation Sequencing for Pathogen Detection. Annu Rev Pathol. 2019 Jan 24;14:319–38.
- 14. Li H, Niu E, Fu J, Huang Y, Gao Y, Chai W, et al. The Clinical Impact of Metagenomic Next-

- Generation Sequencing for the Diagnosis of Periprosthetic Joint Infection. Infect Drug Resist. 2023;16:6521–33.
- 15. Mei J, Hu H, Zhu S, Ding H, Huang Z, Li W, et al. Diagnostic Role of mNGS in Polymicrobial Periprosthetic Joint Infection. J Clin Med. 2023 Mar 1;12(5).
- 16. Higgins E, Suh GA, Tande AJ. Enhancing Diagnostics in Orthopedic Infections. J Clin Microbiol. 2022 Jun 15;60(6).
- 17. Lüdemann M, Sulastyanto S, Raab P, Schoen C, Rudert M. Periprosthetic joint infection: Comparison of automated multiplex-PCR Unyvero i60 ITI cartridge system with bacterial culture and real-time PCR. Technol Health Care. 2022;30(4):1005–15.
- 18. Lausmann C, Kolle KN, Citak M, Abdelaziz H, Schulmeyer J, Delgado GD, et al. How reliable is the next generation of multiplex-PCR for diagnosing prosthetic joint infection compared to the MSIS criteria? Still missing the ideal test. HIP International. 2020 Sep 1;30(1 suppl):72–7.
- 19. Gardete-Hartmann S, Mitterer JA, Sebastian S, Frank BJH, Simon S, Huber S, et al. The role of BioFire Joint Infection Panel in diagnosing periprosthetic hip and knee joint infections in patients with unclear conventional microbiological results. Bone Joint Res. 2024 Jul 10;13(7):353–61.
- 20. Lausmann C, Zahar A, Citak M, Brañes J, Schmidl S, Frommelt L, et al. Are There Benefits In Early Diagnosis Of Prosthetic Joint Infection With Multiplex Polymerase Chain Reaction? J Bone Jt Infect. 2017;2(4):175–83.
- 21. Kato H, Hagihara M, Asai N, Umemura T, Hirai J, Yamagishi Y, et al. Comparison of microbial detection rates in microbial culture methods versus next-generation sequencing in patients with prosthetic joint infection: a systematic review and meta-analysis. J Orthop Surg Res. 2023 Dec 1;18(1).
- 22. Kildow BJ, Ryan SP, Danilkowicz R, Lazarides AL, Penrose C, Bolognesi MP, et al. Next-generation sequencing not superior to culture in periprosthetic joint infection diagnosis. Bone Joint J. 2021 Jan;103-B(1):26–31.
- 23. Hantouly AT, Alzobi O, Toubasi AA, Zikria B, Al Dosari MAA, Ahmed G. Higher sensitivity and accuracy of synovial next-generation sequencing in comparison to culture in diagnosing periprosthetic joint infection: a systematic review and meta-analysis. Knee Surgery, Sports Traumatology, Arthroscopy. 2023 Sep 1;31(9):3672–83.
- 24. Goswami K, Clarkson S, Phillips CD, Dennis DA, Klatt BA, O'Malley MJ, et al. An Enhanced Understanding of Culture-Negative Periprosthetic Joint Infection with Next-Generation Sequencing: A Multicenter Study. Journal of Bone and Joint Surgery. 2022 Sep 7;104(17):1523–9.
- 25. Wouthuyzen-Bakker M, Benito N, Soriano A. The Effect of Preoperative Antimicrobial Prophylaxis on Intraoperative Culture Results in Patients with a Suspected or Confirmed Prosthetic Joint Infection: a Systematic Review. J Clin Microbiol. 2017 Sep;55(9):2765–74.
- 26. Gude S, Pinçe E, Taute KM, Seinen AB, Shimizu TS, Tans SJ. Bacterial coexistence driven by

- motility and spatial competition. Nature. 2020 Feb 27;578(7796):588-92.
- 27. Chotanaphuti T, Courtney PM, Fram B, In den Kleef NJ, Kim TK, Kuo FC, et al. Hip and Knee Section, Treatment, Algorithm: Proceedings of International Consensus on Orthopedic Infections. J Arthroplasty. 2019 Feb;34(2): S393–7.
- 28. Budhiparama NC, Santoso A, Hidayat H, Ifran NN. DAIR (Debridement, Antibiotics, and Implant Retention) for the Treatment of Periprosthetic Joint Infection of Knee. In: Infection in Knee Replacement. Cham: Springer International Publishing; 2022. p. 159–70.
- 29. Darwich A, Dally FJ, Abu Olba K, Mohs E, Gravius S, Hetjens S, et al. Superinfection with Difficult-to-Treat Pathogens Significantly Reduces the Outcome of Periprosthetic Joint Infections. Antibiotics. 2021 Sep 23;10(10):1145.
- 30. Goswami K, Parvizi J. Culture-negative periprosthetic joint infection: is there a diagnostic role for next-generation sequencing? Vol. 20, Expert Review of Molecular Diagnostics. Taylor and Francis Ltd; 2020. p. 269–72.
- 31. Li C, Renz N, Trampuz A, Ojeda-Thies C. Twenty common errors in the diagnosis and treatment of periprosthetic joint infection. Int Orthop. 2020 Jan 22;44(1):3–14.